Acarina 28 (1): 65–74 © Acarina 2020

# REDESCRIPTION OF *GAEOLAELAPS DEBILIS* (MA) (ACARI: MESOSTIGMATA: LAELAPIDAE), WITH A KEY TO WORLD SPECIES OF *GAEOLAELAPS* WITH SETAE *ST1* OFF STERNAL SHIELD

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ABSTRACT: Ma (1996) described *Gaeolaelaps debilis* as a new species from China, being just one female collected from the nest of Daurian ground squirrel, *Spermophilus dauricus Brandt* (Rodentia: Sciuridae), in the Jilin Province. Then he presented a supplementary description of this species, based on female and deutonymph specimens collected from soil in same province from China (Ma 2004); but they both lack some of the most important details concerning leg chaetotaxy and do not provide enough information for accurate and consistent species identification. In this paper, we redescribe the adult female of *G. debilis* (Ma, 1996) based on the newly collected specimens from new localities in China, Iran and the Russian Far East. Our work includes the first description of the male of this species. Moreover, a key to the species of *Gaeolaelaps* with setae *st1* off sternal shield is presented.

KEY WORDS: Phoretic mites, Gamasina, Dermanyssoidea, Hypoaspidinae, taxonomy, Palearctic region.

DOI: 10.21684/0132-8077-2020-28-1-65-74

#### INTRODUCTION

Within the Hypoaspidinae subfamily (Laelapidae family), Gaeolaelaps is one of the largest genera. Although it is better known as a group of predatory mites found in soil and litter (Bregetova 1977; Karg 1993; Beaulieu 2009), sometimes, Gaeolaelaps are recovered from vertebrate (Tenorio 1982) or arthropod nests (Rosario 1981; Strong and Halliday 1994; Fain et al. 1995; Trach 2012, 2016). A few species of this genus have been collected in association with ants (Zeman 1982; Beaulieu 2009; Walter and Moser 2010; Joharchi et al. 2019d, e). More recently, species of Gaeolaelaps have been recorded from beetles. For example, Sklyar (2012) described G. dubininae in association with Amara majuscule (Chaudoir) (Coleoptera: Carabidae). Trach (2012, 2016) described G. carabidophilus associated with Stenolophus mixtus (Herbst), G. khaustovi associated with Bembidion sp. (Coleoptera, Carabidae), as well as Gaeolaelaps heteroceri and G. sevastianovi associated with heterocerid beetles. Joharchi and Babaeian (2014) described G. saboorii from under the elytra of Acinopus sp. in Iran. Trach and Joharchi (2018), in their review of Laelapidae mites in association with scarab beetles in Ukraine, recorded Gaeolaelaps rhizotrogi (Mašan) on Amphimallon solstitiale (L.) (Coleoptera, Scarabaeidae) for the first time in this country. Finally, Saeidi et al. (2019) described Gaeolaelaps scarites Joharchi and Saeidi recovered from under the elytra of Scarites (Parallelomorphus) terricola Bonelli (Coleoptera: Carabidae) in Iran.

During the survey of free-living and insect-associated species of laelapid mites in China, Iran and Russia, a series of female and male specimens identified as G. debilis (Ma, 1996) were collected from different habitats. Ma (1996, 2004) considered G. debilis as a member of very broadly conceived genus Hypoaspis sensu lato. Subsequently, Beaulieu (2009) treated this species as a member of Gaeolaelaps. The descriptions provided by Ma (1996, 2004) lack some of the most important details concerning leg chaetotaxy and do not provide enough information for accurate and consistent species identification. Therefore, Gaeolaelaps debilis has never been fully described. Towards this aim, we herein provide additional morphological characters to complement the previous descriptions (Ma 1996, 2004) of G. debilis on the basis of adult females, collected from the following contexts: soil in Shanghai, China (close to type locality in the Jilin Province, China); the nest of Formica sp., in the vicinity of Khabarovsk (Russian Far East); and under the elytra of Cetonia sp. (Scarabaeidae) in the Yazd Province, Iran. Our work also includes the first description of the male of this species.

Moreover, we provide a world key to the species of *Gaeolaelaps* with setae *st1* off sternal shield.

# **MATERIALS AND METHODS**

The host beetles were collected by hand, using light traps, after which they were transferred into vials containing 70% ethyl alcohol. The host ants were collected from a forest nest using an aspirator and

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placed in vials with 70% ethanol. Thereafter, alcohol sediments from the vials were inspected for detached phoretic mites. The mites were extracted from soil samples using Berlese–Tullgren funnels. The mites were cleared in a lactic acid solution and mounted in Hoyer's medium (Walter and Krantz 2009).

The line drawings and examinations of the specimens were performed using the Zeiss Axio Imager A2 and the Leica DM 2500 compound microscopes, equipped with differential interference contrast and phase contrast optical systems, attached to the AxioCam ICc 5 and ICC50 HD cameras, respectively. The figures were elaborated with Adobe Photoshop CS2 software, based on line drawings. The images and morphological measurements were taken with the help of ZEN 2012 (v. 8.0) and Leica Application Suite (LAS) (v. 4.2, Live and Interactive Measurements modules) software. The photomicrographs were taken with an AxioCam 506 camera (Carl Zeiss, Germany).

The measurements of structures are expressed as ranges (minimum-maximum) in micrometers (µm). The length and width of the dorsal shield were taken from the anterior to posterior margins along the midline, and at the level of r3, respectively. The length and width of the sternal shield were measured at the maximum length and at the broadest points (at level of endopodal between coxae II and III), respectively. The length of the genital shield was measured along the midline from the anterior margin of the hyaline extension to the posterior margin of the shield, and its width where maximal, posteriorly to genital setae st5. The leg length was measured from the base of the coxa to the apex of the tarsus (excluding the pre-tarsus). The nomenclature used for the dorsal idiosomal chaetotaxy follows that of Lindquist and Evans (1965). The notations for leg and palp setae follow those of Evans (1963a, b). Other anatomical structures mostly follow Evans and Till (1979). Notations for the idiosomal porelike structures (gland pores and poroids/lyrifissures) and for the peritrematal shield follow mostly Athias-Henriot (1971, 1975). The notations for the porelike structures on the sternal shield and for the peritrematal shield region also follow modifications and additions by Johnston and Moraza (1991), adapted by Kazemi et al. (2014).

### **SYSTEMATICS**

Family **Laelapidae** Genus *Gaeolaelaps* **Evans and Till, 1966** *Hypoaspis* (*Gaeolaelaps*) Evans and Till 1966: 159. Type species: *Laelaps aculeifer* Canestrini, 1884, by original designation.

**Diagnosis**. The concept of *Gaeolaelaps* used here is based on that of Beaulieu (2009) with modifications by Kazemi *et al.* (2014).

### Gaeolaelaps debilis (Ma, 1996)

(Figs. 1–4)

Hypoaspis debilis Ma, 1996: 51. Hypoaspis debilis.—Ma 2004: 83, Ren and Guo 2008: 329.

Gaeolaelaps debilis.—Beaulieu 2009: 37, Kazemi et al. 2014: 504, Moreira 2014: 247, Yan et al. 2018: 261.

Diagnosis (female). Dorsal shield oval-shaped, with weak reticulation, more distinct in opisthonotal and lateral regions, bearing 39 pairs of setae (21–31), 22 pairs of podonotal and 17 pairs of opisthonotal setae, including two pairs of Zx setae, two supernumerary setae Jx present between J-series. All setae relatively short, never reaching base of next posterior setae, except j1, J5, Z5. Presternal area lightly sclerotized, with a few transverse curved lines, sternal shield smooth almost throughout, except a longitudinally lateral part between iv1 and st3 faintly reticulated with curved lines, bearing two pairs of smooth pointed setae, st1 off shield, on poorly sclerotized cuticle, anterior margin slightly convex, ratio of shield length/width (at broadest level)  $\approx 0.81$ . Genital shield elongated, ratio of length/width (at broadest level) ≈2.2, surface without any distinct reticulation, almost completely smooth. Anal shield longer than wide, with length/width ratio (at broadest level)  $\simeq 1.16$ , gland pores gv3 located on soft opisthogastric cuticle close to shield. Peritremes long, reaching to slightly beyond mid-level of coxae I. Palp tarsal apotele two-tined, fixed digit of chelicera with six teeth. Tarsus IV without elongate setae.

# **Redescription**. *Female* (n=11 specimens).

Dorsal idiosoma (Figs. 1A, 2A). Dorsal shield oval-shaped, 539–575 long, 288–315 wide, covering entire idiosoma, with weak reticulation, more distinct in opisthonotal and lateral regions. Shield bearing 39 pairs of setae, 22 pairs of podonotal and 17 pairs of opisthonotal setae, including two pairs of Zx setae, two supernumerary setae Jx present between J-series. All setae relatively short (21–31), never reaching base of next posterior setae, except j1, J5 (30–32), Z5 (44–46). Shield with 21 pairs of discernible pore-like structures, including 16 poroids (id1, id2, id4, id5, idm1–idm6, idx, is1, idl1,

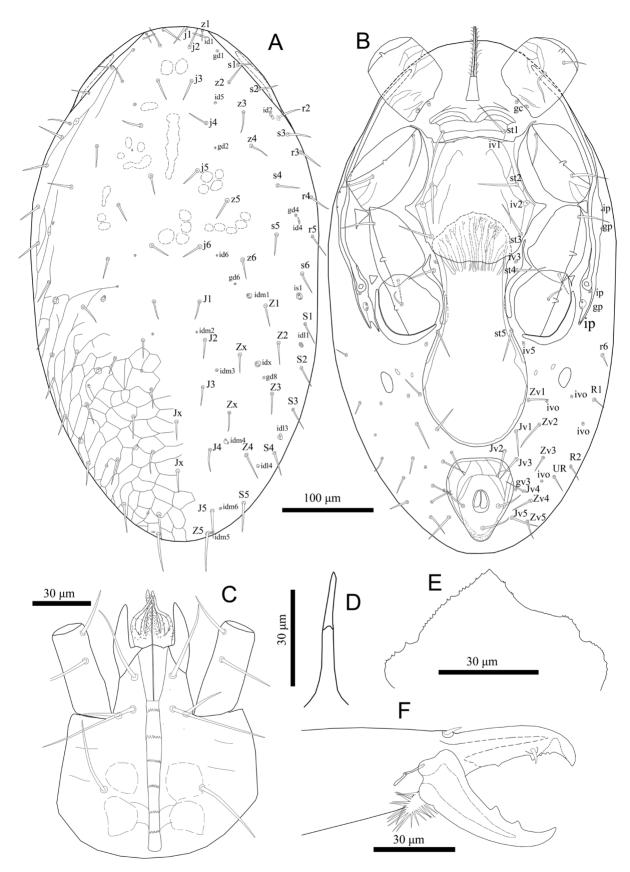


Fig. 1. *Gaeolaelaps debilis* (Ma, 1996), female. A—dorsal idiosoma; B—ventral idiosoma; C—subcapitulum; D—supralabral process; E—epistome; F—chelicera.

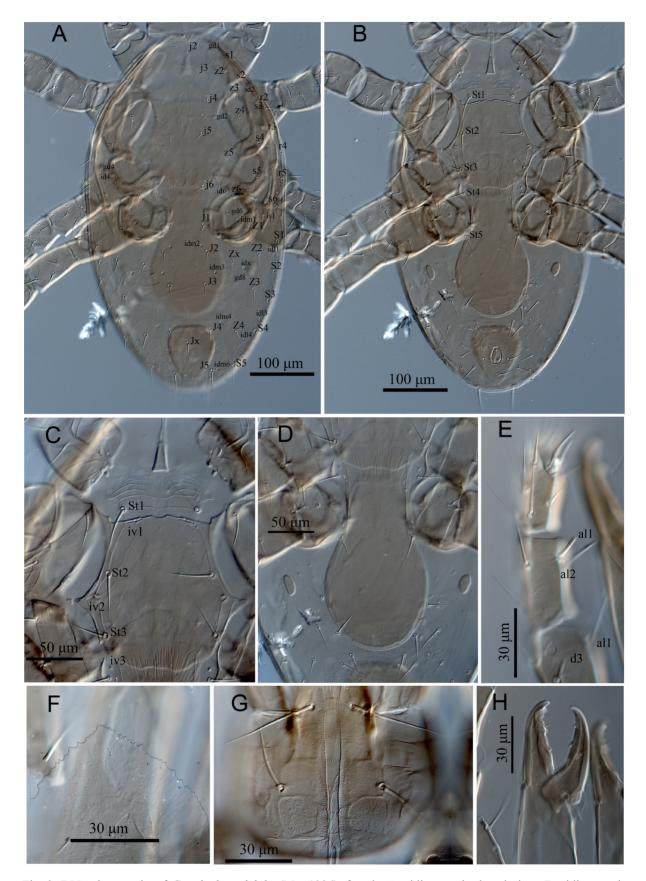
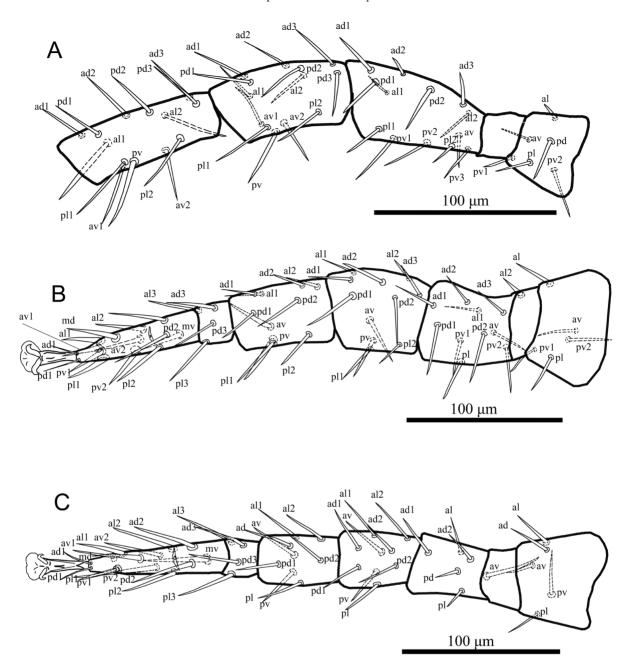


Fig. 2. DIC micrographs of *Gaeolaelaps debilis* (Ma, 1996), female. A—idiosoma in dorsal view; B—idiosoma in ventral view; C—sternal shield; D—genital shield and opisthogastric area; E— distal portion of palp, with a focus on apotele; F—epistome; G—hypostome; H—chelicera.



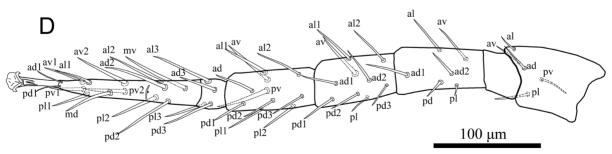


Fig. 3. Gaeolaelaps debilis (Ma, 1996), female. A—leg I (trochanter-tibia); B—leg II; C—leg III; D—leg IV.

idl3, idl4) and five gland openings (gd1-2, gd4, gd5, gd8) (Figs. 1A, 2A).

*Ventral idiosoma* (Figs. 1B, 2B–D). Tritosternum with paired pilose laciniae (74–77), columnar

base 26–28×13–16 wide; presternal area lightly sclerotized, with a few transverse curved lines. Sternal shield (length 126–136) narrowest between coxae II (95–102), widest (158–167), with anterior

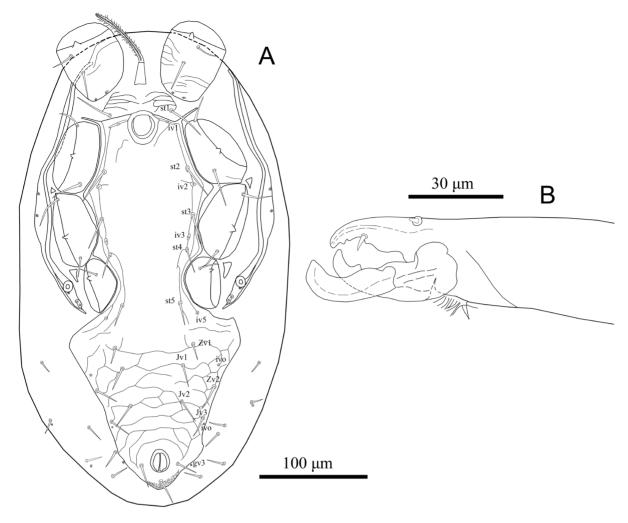


Fig. 4. Gaeolaelaps debilis (Ma, 1996), male. A—dorsal idiosoma; B—chelicera.

margin slightly convex, anterolateral corners of shield extending between acetabula I-II; shield bearing two pairs of smooth setae (st2 40–42, st3 38–40), st1 (42–44) off shield, on poorly sclerotized cuticle (Fig. 2 B, C), setae not reaching base of next setae, one pair of lyrifissures adjacent to setae st1, and a pair of poroids between st2 and st3; sternal shield smooth almost throughout, except a longitudinally lateral part between iv1 and st3 faintly reticulated with some curved lines. Metasternal setae st4 (33-36) and metasternal poroids located on soft integument; metasternal platelets absent. Endopodal plates II/III completely fused to sternal shield, endopodal plates III/IV elongate, narrow, and curved. Genital shield elongated, slightly expanded laterally past level of setae st5, length 245-263, maximum width 108-118, posterior margin rounded, without any distinct reticulation, almost completely smooth (Fig. 2D); bearing a pair of simple setae st5 (28–30); paragenital poroids iv5 located on soft cuticle lateral to shield near seta st5

(Fig. 2D), clearly separated from anal shield, the distance midline between posterior margin of genital shield and anterior margin of anal shield nearly as long as anal opening. Anal shield subtriangular, rounded anteriorly, longer than wide, length 85-93, width 75-80, anterior half lineatereticulate, para-anal setae (27–29) shorter than post-anal seta (30–32), cribrum small, with 3–4 irregular rows of spicules, mostly limited to region posterior to post-anal seta, pair of anterior arms reaching about to midlevel of distance between post-anal and para-anal setae; gland pores gv3 located on soft opisthogastric cuticle close to shield. Soft opisthogastric cuticle with two pairs of small, oval paragenital platelets between st5 and ZV1 and pair of oval metapodal plates (22–24 long×8–13 wide), 14 pairs of smooth setae (19–33) (Jv1–Jv5, Zv1-Zv5, R1, R2, UR, r6), five pairs of poroids (iv5 and four pairs of ivo) and gland pores gv3. Two small and subtriangular exopodal platelets between coxae II-III and III-IV present; parapodal platelets

strip-like (Fig. 1B). Peritreme extending anteriorly to slightly beyond mid-level of coxa I; peritrematal shield narrow, fused to dorsal shield near setae s1, bearing one pair of gland pores (gp) near external margin of shield at level of anterior edge of coxae III and one pair of poroids (ip) at level of posterior edge of coxae II, free from exopodal shields; post-stigmatic region with a longitudinal line from stigmata to shield apex, and with two lyrifissures ip and a gland pore gp (Fig. 1B).

Gnathosoma (Figs. 1C-F and 2E-H). Epistome subtriangular, more or less pointed apically, anterior margin finely denticulate (Figs. 1E, 2F). Hypostomal groove with six rows of denticles, each row with 7-13 denticles, with smooth anterior and posterior transverse lines (Figs. 1C, 2G). Hypostome with four pairs of setae, internal posterior hypostomal setae h3 (50-52), h1 (42-44), h2 (22-24), palpcoxal pc (38-40) (Figs. 1C, 2G). Internal malae fringed, slightly longer than corniculi, with a pair of long median projections and two additional pairs of thinner, shorter lateral projections, the inner pair shortest (Fig. 1C). Corniculi robust and horn-like, extending slightly beyond palptrochanter. Supralabral process with a narrow and undivided projection (Fig. 1D). Chaetotaxy of palps: trochanter 2, femur 5, genu 6, tibia 14, tarsus 15, all setae smooth and needle-like except al1, al2 on palp genu thickened and al1 apically spatulate, palp tarsal apotele two-tined (Fig. 2E). Fixed digit of chelicera with an offset and most distal tooth (gabelzhan), followed by 4-5 variously sized teeth, a setaceous pilus dentilis, dorsal cheliceral seta prostrate, arthrodial membrane with a rounded flap and normal filaments; cheliceral lyrifissures distinct, movable digit with two teeth (Figs. 1F, 2H).

Legs (Fig. 3A–D). Legs II (377–406) and III, (329–382) short, I (497–504) and IV (540–548) longer. Chaetotaxy normal for free-living Laelapidae: Leg I (Fig. 3A): coxa 0-0/1, 0/1-0, trochanter 1-0/1, 1/2-1 (pd slightly thickened), femur 2-3/1, 2/3-2 (pd2 slightly thickened), genu 2-3/2, 3/1-2, tibia 2-3/2, 3/1-2 (pv and pl1 slightly thickened). Leg II (Fig. 3B): coxa 0-0/1, 0/1-0, trochanter 1-0/1, 0/2-1, femur 2-3/1, 2/2-1 (al2 thickened), genu 2-3/1, 2/1-2 (all ventral setae thickened). Leg III (Fig. 3C): coxa 0-0/1, 0/1-0, trochanter 1-1/1, 0/1-1, femur 1-2/1, 1/0-1, genu 2-2/1, 2/1-1 (all ventral setae slightly thickened), tibia: 2-1/1, 2/1-1 (all ventral setae slightly thickened). Leg IV (Fig. 3D):

coxa 0-0/1, 0/0-0, trochanter 1-1/1, 0/1-1, femur 1-2/1, 1/0-1 (*ad1* longest), genu 2-2/1, 3/0-1 (*av* slightly thickened), tibia 2-1/1, 3/1-2 (all ventral setae thickened). Tarsi II–IV with 18 setae (3-3/2, 3/2-3 + *mv*, *md*); tarsus II–IV with some spine-like ventral and lateral setae (Fig. 3B–D), and tarsus IV without elongate setae. All pretarsi with well-developed paired claws, rounded pulvilli and normal ambulacral stalk.

# **Description**. *Male* (n=3 specimens)

*Dorsal idiosoma*. Dorsal shield (445–450) long, (220–228) wide; ornamentation and chaetotaxy as in female.

Ventral idiosoma (Fig. 4A). Sternal, genital, endopodal, ventral and anal shields fused into holoventral shield, weakly reticulate, distinctly posteriorly to Zv1 level, bearing st2–5, st1 off shield, on poorly sclerotized cuticle, five pairs of opisthogastric setae (Jv1–Jv3, Zv2, Zv3) in addition to circumanal setae; gland pores gv3 on soft opisthogastric cuticle, close to para-anal setae, gland pore gv2 behind coxa IV not discerned; cribrum with 3–4 irregular rows of spicules, anterior arms reaching about to mid-level of distance between post-anal and para-anal setae (Fig. 4A). Soft opisthogastric cuticle with six pairs of smooth setae, two pairs of poroids and gland pores gv3.

Gnathosoma. Epistome and subcapitulum similar to female. Fixed digit with two various size of teeth, a setaceous pilus dentilis. Movable digit of chelicera with one median tooth, spermatodactyl relatively stout (0.65 × as long as movable digit) and curved dorsally, with bluntly pointed tip; dorsal seta conspicuous, markedly reduced, fringed hyaline arthrodial process at base of movable digit (Fig. 4B). Palps similar to those of female.

Legs. Chaetotaxy as in female.

Specimens examined and deposition: China, five females, three males, Shanghai, Gongqing Forest Park, 31°19′N, 121°32′E, 28 May 2017, O. Joharchi coll., from soil—deposited in the collection of the Tyumen State University Museum of Zoology, Tyumen, Russia; Iran, two females, Yazd Province, Manshad Village, 31°30′N, 54°13′E, 19 May 2016, P. Kafi coll., under the elytra of *Cetonia* sp. (Scarabaeidae)—deposited in the Acarological Collection, Faculty of Agriculture, Shiraz Branch, Islamic Azad University; Russia, four females, vicinity of Khabarovsk, 48°14′N 135°01′E, 15 August 2018, A.A. Khaustov coll., in the nest of *Formica* sp.—deposited in the collection of the Tyumen State University Museum of Zoology, Tyumen, Russia.

Remarks. Although the diagnosis above is based primarily on specimens from Shanghai, China (close to the type locality in Daan, Jilin Province, China), it is supplemented by comparisons with the specimens collected from Iran and the Russian Far East, as well as with the descriptions provided by Ma (1996, 2004). Although our efforts to view type material were unsuccessful, our specimens are in perfect agreement with the incomplete descriptions and illustrations by Ma (1996, 2004). Gaeolaelaps debilis has the unusual character state of setae st1 off sternal shield. Sternal shield in *Gaeolaelaps* usually bears three pairs of setae, rarely setae st1 off shield in the presternal region. This occurs only in eight species of Gaeolaelaps (Beaulieu 2009; Kazemi et al. 2014; Joharchi et al. 2019a-c). However, seta st1 being off the sternal shield in the presternal region can be difficult to discern. In particular, in some of the aforementioned species, seta st1 could be borne on paired anterior extensions of the shield, as in G. jondishapouri (e.g., Kazemi et al. 2014). The following key is based on direct examination of specimens, except for G. krantzi, G. franzi and G. genitotortus, in which cases we have relied on previously published descriptions and illustrations.

# Key to species of Gaeolaelaps with setae st1 off sternal shield

plates III/IV obviously free from sternal shield, surface of sternal and genital shields smooth almost throughout......6 6. Dorsal shield obviously narrowing from level of setae r3, progressively tapering, genital shield not extending close to anal shield, the distance midline between posterior margin of genital shield and anterior margin of anal shield more than twice as long as anal opening ..... G. aculeiferoides (Teng, 1982) — Dorsal shield oval-shaped, genital shield extending close to anal shield, the distance midline between posterior margin of genital shield and anterior margin of anal shield nearly as long as anal opening......7 7. Size of body small (390 long, 180 wide) and leg I longer than idiosoma, anal shield nearly wider than long, post-stigmatic section elongated, slender and defectively developed after stigma..... — Size of body much larger (539–575 long, 288– 315 wide) and leg I shorter than idiosoma, anal shield longer than wide, post-stigmatic section well devel-

Gaeolaelaps debilis was originally described from Daan, Jilin Province, China on the basis of only one female specimen collected from the nest of Spermophilus dauricus Brandt (Rodentia: Sciuridae) (Ma 1996). Then he collected this species from soil in the same province in China (Ma 2004). In our study, a few G. debilis specimens have been collected from the nest of Formica sp. in the Russian Far East, as well as from under the elytra of Cetonia sp. in Iran. All the examined phoretic individuals were adult females; no males or immatures were collected. The collected specimens of G. debilis have normal morphology, like other free-living Gaeolaelaps species (e.g., dentate chelicerae; sclerotized, horn-like corniculi, etc.), and none of the phoretic individuals were swollen, which points to it being unlikely that G. debilis is parasitic.

It is not possible to draw any firm conclusions about *G. debilis*'s host specificity but its normal morphology suggests that it may have a mutualistic symbiosis relationship with its hosts. In particular, this mite may feed on other small invertebrates, such as immature stages of Astigmata mites living in ant colonies, in microhabitats created by beetles, or in the nests of rodents. In the soil, *G. debilis* is probably a predator of small soil invertebrates, like other representatives of the *Gaeolaelaps* genus. We stress that further experimental work is needed to establish the true role of this mite in

relation to its insect and rodent hosts, as well as to the soil ecosystem.

### **ACKNOWLEDGEMENTS**

This paper is part of a Ph.D. dissertation by the senior author, which was financially supported by the Department of Entomology, Faculty of Agriculture, Shiraz Branch, Islamic Azad University, Shiraz, Iran. We cordially thank A.A. Khaustov (Tyumen State University, Tyumen, Russia) for making the materials from the Russian Far East available. The research of the corresponding author (Omid Joharchi) was supported by a project sponsored by the Russian Science Foundation (No. 20-64-47015). We are grateful to the anonymous reviewers for their valuable suggestions.

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